5

10

15

Leptin antagonist and method for quantitative measurement of leptin

20

This invention relates to specific antibodies or fusion proteins, in particular a specific antibody A or a fusion protein directed to a leptin receptor (leptin-R) or a leptin-binding protein (leptin-BP), as well as to the use of these antibodies or fusion proteins for quantitative analysis, for therapeutic purposes and for the preparation of

25　therapeutic drugs. Furthermore, the invention relates to a method for quantitative determination of leptin in the sample of solubilized or suspended leptin-binding proteins by using specific antibodies or fusion proteins according to the invention, as well as to diagnostic agents and (diagnostic) kits containing this antibody or fusion protein.

30

Leptin (derived of the Greek term leptos = thin) is a protein hormone, that is primarily segregated by the fat cells (adipocytes). In 1994, leptin (Zhang et al. (1994) Na-

2

ture 372, 425) was discovered as genetic product of the obesitas gene (ob gene) with a molecular weight of approx. 16 kDa, formed by 146 amino acids. It plays an important role in energy metabolism (Friedman et al., (1998) Nature 395, 763 – 770). Apart from its relevance for energy metabolism, its contribution to the modula-

5   tion of immunocompetent cells (Lord et al., (1998) Nature 394, 6696) or haemoto-poietic cells (Sierra-Honigmann et al., (1998) Science 281, 1683-1686) has been described in the meantime as well as a permissive function in the induction of pu-berty (Quinton et al., (1999) J Clin Endocrinol Metab 84(7), 2336-41). Leptin has also been strongly associated with diabetes and chronic heart failure (CHF) (see E.M.

10   El-Bindary and A.Z. Darwish, Volume 7, Nos 4/5, July - September 2001, 697- 706).

In mice (*ob/ob* mouse*)*, the lack of the leptin gene (*ob/ob* mouse*)* leads to massive overweight (adipositas). Due to the fact, that administration of leptin to *ob/ob* mice induces reduced food uptake and finally weight reduction, leptin was attributed

15   some importance as an appetite suppressant. Anyhow, the situation is far more complex.

The effect of leptin is mediated via the leptin receptor (leptin-R) (Tartaglia et al., (1995) Cell 83(7), 1263-71), thus activating intracellular signal cascades. The leptin

20   receptor pertains to the so-called class I of the cytokine receptor superfamily. In many receptors of this family, an extra-cellular portion of the leptin receptors circu-lates as leptin-binding protein in the blood, and this is also the case for the leptin receptor.

25   Defects of the leptin receptor were identified as a cause of overweight problems in human beings (Clement et al., (1998) Nature 392, 398 – 401). Conversely, patients suffering from anorexia nervosa (diminished appetite) seemingly have increased leptin levels in relation to their reduced fat mass. Therefore, determination of leptin concentration in blood or in serum samples is an important diagnostic tool for clari-

30   fication of the underlying cause of eating disorders or extreme obesity.

3

As mentioned above, El-Bindary and Darwish (2001) have strongly associated leptin with diabetes and chronic heart failure (CHF). El-Bindary and Darwish (2001) investigated the interaction of TNF-α, leptin levels and insulin in patients suffering from

5   CHF. CHF constitutes a complex syndrome associated with disturbance in several metabolic and endocrine functions. There is accumulated evidence that weight loss and cachexia seen in advanced stages of CHF constitute a poor prognosis in patients suffering therefrom. In particular, an increased plasma leptin level is assumed to be implicated in the wasting associated with late stage heart failure. Additionally,

10   chronic heart failure is characterized by a hyperinsulinaemic state (Swan et al., Journal of the American College of Cardiology, 1997, 30:527–32), in which there is striking loss of both muscle and adipose tissue leading to overt cardiac cachexia in these patients (see Cleland and Clark, European heart journal, 1998, 19:1421–2). Recent attention has therefore been focused on cytokine activity in CHF. According

15   to El-Bindary and Darwish (2001), there is a significant elevation of TNF-α in late stage CHF. Apparently, the level of circulating TNF-α is elevated in patients with advanced CHF. Assumably, TNF-α may contribute to the progression of the cardiac decompensation that occurs in advanced cases of CHF. The results also reveal a correlation between TNF-α and plasma leptin concentration. TNF-α may act di-

20   rectly on adipocytes to increase the production of the lipostatic factor leptin, wherein the level of serum leptin appears to be under control of TNF-α. Hence, there is a significant elevation of the insulin level and a correlation with TNF-α in late stage CHF. El-Bindary and Darwish (2001) also found evidence for a positive correlation between plasma leptin and insulin level in late-stage disease. Increased

25   plasma leptin and associated increase in insulin were considered to be another cachexia causing factor. Increase in plasma TNF-α , leptin and insulin levels, and positive correlation between them in late stage heart failure may constitute one of many vicious circles of advanced stage CHF.

4

Since disorders of energy metabolism, in particular eating disorders, such as ano-rexia nervosa (diminished appetite), are observed more frequently, the identification of effective leptin-antagonists that may reduce, inhibit or even block the function of leptin, is of major interest and economic significance.

5

Therefore, it is a first object of the present invention to provide an antagonist of leptin-R and leptin-BP. Such an antagonist may be suitable in the treatment of dis-eases and/or symptoms associated with excessive leptin levels, such as diabetes, cachexia, obesity, chronic heart failure (CHF), etc., as well in the development and

10   provision of a medicament and/or therapies for such diseases.

By using suitable leptin-antagonists (as mentioned above), regulatory mechanisms and specific effects of leptin are amenable to examination *in vitro* by selective inhi-bition of the leptin receptor and/or displacement of leptin from leptin-binding pro-

15   teins.

In the light of the above, the quantitative determination of leptin in a sample and in particular in physiological body liquids – such as e.g. blood - is a prerequisite for therapeutic treatment and an important diagnostic tool. Many e.g. hormones or

20   other messenger substances (ligands), bind specifically to their membrane receptors, the extracellular domain(s) of which is (are) frequently solubilized or suspended in the liquid. Thereby, a complex composed of ligands to be determined, e.g. leptin (ligand) and the leptin-binding protein, is formed, which influences (reduces) the level of free ligand in the sample. In order to avoid that problem, a labeled ligand is

25   used and added to the sample. Nevertheless, the binding protein may bind to the labeled ligand in the solution and thus, the ligand is not detectable in the sample. Therefore, the binding protein interferes with quantitative ligand measurement and diagnostics.

5

Generally, so-called immunoassays are used for the analysis of the leptin concentration. They are based on the principle of an interaction of specific antibodies with an analyte. Alternatively, competitive assays are used (such as the radioimmunoassay, RIA), in which labeled leptin competes with the leptin present in the sample for the

5   binding site on an antibody, thus generating a signal being inversely proportional to the concentration of the analyte to be determined. However, nowadays, sandwich-immunoassays are used most frequently, in which an immobilized specific antibody binds the analyte ("capturing antibody"), and subsequently, a second, labeled antibody, directed to a different epitope of the analyte, binds the analyte, which is then

10   immobilized. This generates a signal, that is proportional to the amount of the bound analyte. The best-known example of this measurement method is the "enzyme linked immuno sorbent assay" (ELISA) with a colorimetric endpoint. Other possibilities for the generation of a signal are radioactivity, chemoluminescence or (time resolved) fluorescence.

15

All assay methods mentioned above are based on the principle of the highly affine, specific binding between an antibody directed to a e.g. hormone ligand – leptin, in this case - and the hormone ligand in the sample to be analyzed.

20   The main source of interference in the measurement of leptin in serum or blood is the presence of a highly affine leptin-binding protein, namely the soluble extracellular portion of the leptin receptor in human serum. The effect of this leptin-binding protein on the measurement result may cause erroneously high or erroneously low values, always depending on the type of assay used (competitive assay or

25 · sandwich-assay). In the competitive assay, on the one hand, the labeled leptin (as a so-called "tracer") may be bound by the leptin-binding protein, thus being removed from solution, resulting in erroneously high concentrations. On the other hand, the leptin-binding protein may also block sterically the interaction of the specific antibody with the leptin molecules from a serum sample and thus may also cause erro-

30   neously low concentrations in the competitive assay. In contrast, this steric obstruc-

6

tion of the interaction between antibody and hormone generally provokes errone-
ously low results in the sandwich-immunoassay, since less hormone or less detec-
tion antibody will be bound. The error in the measurement results caused by this
interference will become particularly relevant in the presence of certain physiologi-
5   cal or pathological conditions that are associated with a change of leptin concentra-
tion in the blood.

Since Leptin-BP binds leptin in a sample and thus reduces the amount of free leptin
in the sample, no unequivocal determination of physiological leptin concentrations
10   can be obtained. Thus, due to the presence of soluble leptin-binding proteins, all
experimental data for body fluid leptin levels are biassed by interferences between
leptin and leptin-binding protein.

In order to develop a standardizable diagnostic method or a therapeutic treatment
15   for diseases such as obesity, diabetes and/or chronic heart failure (CHF), etc., there
is a need for selective, specific and effective substances or molecules that bind to a
leptin receptor and/or a leptin-binding protein. To date, only few substances or
molecules are known, which selectively, specifically and effectively bind to a leptin
receptor and/or a leptin-binding protein. Selectivity and specificity are prerequisites
20   for quantitative and qualitative determination. Gonzales et al. (Gonzales et al.,
(2003) Mol Hum Reprod 9(3), 151-8) for instance report on polyclonal antibodies,
that seem to inhibit leptin effects to a certain extent. However, such polyclonal anti-
bodies cannot be reproduced, they cannot be humanized and are available only in
limited amounts.

25

On the one hand, alternative solutions to avoid measurement inaccuracies relate to
methodic extraction methods, that are carried out before the actual measurement
method itself, in order to eliminate, e.g. the leptin-binding proteins. Anyhow, this
implies significantly increased efforts as to the methods used and may simultane-
30   ously lead to misleading results. Altogether, to date there is no satisfactory state-of-

7

the-art-solution to the difficulties of eliminating soluble leptin-binding proteins in samples to be analyzed.

Therefore, it is a second object of the invention to provide a method to obtain reli-
5   able results of the ligand level, e.g. leptin, of a sample containing leptin-binding protein(s) or leptin receptors with its /their ligand(s):

This object is solved by the present invention, which may render test methods, in particular assays for (body liquid) leptin determination, less susceptible to falsifying
10 interference by leptin-binding protein. Such method may i.a. be suitable for the diagnosis of diseases and/or symptoms associated with excessive leptin levels, such as diabetes, cachexia, obesity, chronic heart failure (CHF), etc..

The inventors of the present invention succeeded in identifying and providing a
15 molecule - in form of an antibody - that in essence specifically prevents the binding of leptin to a leptin receptor (leptin-R) and displaces leptin, which is bound to a leptin-binding protein, from this leptin-binding protein (leptin-BP).

Therefore, the invention provides an antibody A against a leptin receptor (leptin-R)
20 and/or a leptin-binding protein (leptin-BP), characterized in that it fundamentally reduces and, preferably, prevents the interaction of the leptin receptor and/or of the leptin-binding protein with its ligand, i.e. leptin.

A "leptin-binding protein" or "binding protein" (both termed leptin-BP) or "leptin
25 receptor" (leptin-R) within the meaning of this invention comprises all proteins, that can bind with high affinity (specifically) to leptin to be determined in a sample. Leptin-binding protein is usually soluble or be present in the sample as a suspension. Leptin-binding protein is usually found in body liquid samples, e.g. serum, as the extra-cellular portion of the leptin receptor. Anyhow, it may also be located
30 close to the cell membrane. Examples for human sequences of such leptin-binding

proteins or binding proteins or leptin receptors are the sequences O95214, P48357 and O15243 (source: Swiss-Prot/TrEMBL). Preferably they are leptin-binding proteins or binding proteins or leptin receptors of human origin, however, this invention also comprises corresponding proteins of all other vertebrates, in particular

5   mammals, such as rat, mouse, pig, horse, cattle. Examples for such sequences are O02671, Q9MYL0, P48356, Q62959, O89013, Q9JLS8 (source: Swiss-Prot/TrEMBL). Preferably, the leptin-binding protein is a physiological leptin-binding protein, solubilized or suspended in liquid, preferably body liquid.

10   According to the present application, the term "antibody" comprises monoclonal antibodies, polyclonal antibodies, particularly polyclonal monospecific antibodies (i.e. antibodies with different variable regions, which however all recognize a specific epitope), as well as chimeric antibodies, (anti-)anti-idiotypic antibodies (directed to the inventive antibodies), and genetically manipulated antibodies that are

15   all present in bound or soluble form and may – if appropriate - be labeled by "markers" (for example fluorescence marker, gold marker, coupled enzymes). The term "antibody" in the meaning of the present invention typically refers to full-length antibodies of the afore mentioned antibodies. A "full-length" (monoclonal) antibody in the meaning of the present application may be any of the above mentioned inven-

20   tive antibodies in its full-length form.   A full-length antibody of the present invention typically comprises both the domains of the heavy chain and the light chain. The heavy chain of the inventive antibody includes domains C_H1, C_H2 or C_H3 of the constant region and the variable heavy (V_H) immunoglobulin domain. The , the light chain of the inventive antibody includes the variable light immunoglobulin

25   domain (V_L) and the constant light immunoglobulin domain (C_L). Antibodies, not containing all the aforementioned domains or regions of an antibody are fragments of antibodies within the meaning of the present invention. Fragments of antibodies according to the present invention are further defined below.

9

Apart from the inventive antibodies as such, antibodies according to this invention may also be provided as portion of a fusion protein containing other (protein)-constituents. Fusion proteins containing an inventive antibody or its fragment thereof are defined below. All of the aforementioned embodiments fall under the scope of an inventive antibody A.

Antibodies according to the present invention may pertain to one of the following immune globulin classes: IgG, IgM, IgE, IgA, GILD and, if applicable, a subclass of the aforementioned classes, such as the subclasses of the IgG or their mixtures. IgG and its subclasses such as IgG1, IgG2, IgG2a, IgG2b, IgG3 or IgGM are preferred. The IgG subtypes IgG1/k or IgG2b/k are specifically preferred.

Antibodies in the sense of this invention are furthermore proteins or possibly other structures produced by vertebrates or by artificial production methods, that bind with high affinity to a determined surface conformation (epitope) of an antigen, i.e. of another molecule, preferably mono- or polyclonal (partial) structures of the above mentioned immune globulins or also polyclonal monospecific antibodies. Typically, such antibodies contain at least the variable part of immune globulins, and, as the case may be, at least one domain of the constant domain of immune globulins, too.

"Polyclonal antibodies" in the meaning of the present application are typically heterogeneous mixtures of antibody molecules, produced from animal serums, that had been immunized with an appropriate antigen, i.e. a ligand of the antibody A of the present invention, preferably with leptin-BP and/or leptin-R.

A "monoclonal antibody" contains a fundamentally homogeneous population of antibodies, that are directed specifically to antigens, and fundamentally, antibodies show similar epitope-binding sites here. The different antibody variants with mono-specificity may belong to the immune globulin classes described above. They may also be mixtures of different major primary classes or subclasses, preferentially, they

10

consist of a homogenous mixture of IgG-antibodies. This homogeneity may also be achieved by an additional purification step (immuno- precipitation, chromatography, for example by using antibodies directed to IgG).

5   Monoclonal antibodies may also be obtained by using methods known in the state-of-the-art (e.g. . Köhler und Milstein, Nature, 256, 495-397, (1975); US-Patent 4,376,110; Ausübel et al., Harlow und Lane "Antikörper": Laboratory Manual, Cold Spring, Harbor Laboratory (1988); Ausubel et al., (eds), 1998, Current Protocols in Molecular Biology, John Wiley & Sons, New York)). The aforementioned references

10   are included herein in their entirety. "Monoclonal" is intended to mean in particular the product of an artificial construct, in which an antibody-producing cell (B-cell) is fused with an immortalized cancer cell (hybridom), creating a hybridoma cell. Specific antibodies, that are all exclusively directed to one epitope, are produced by this cell. A hybridoma-cell clone, producing monoclonal antibodies according to

15   the present invention, is cultured *in vitro*.

"Genetically manipulated antibodies" according to the present invention may also be produced using the methods described in the aforementioned publications.

20   "Chimeric antibodies" according to the present invention are molecules, that contain different constituents, which are derived from different animal species (e.g. antibodies, showing a variable region, that is derived from a monoclonal mouse antibody and a constant region of a human immunoglobulin). Chimeric antibodies are preferably used on the one hand for the reduction of immunogenicity, if adminis-

25   tered, and on the other hand for the increase of yield, e.g., murine monoclonal antibodies yield higher rates of production from hybridoma cell lines, however, they are also associated with a higher immunogenicity in humans. Therefore, human/murine chimeric antibodies are preferably used. Chimeric antibodies and methods for their production are known state-of-the-art methods. (Cabilly et al., Proc. Natl. Sci. USA

30   81: 3273-3277 (1984); Morrison et al. Proc. Natl. Acad. Sci USA 81:6851-6855

(1984); Boulianne et al. Nature 312 643-646 (1984); Cabilly et al., EP-A-125023; Neuberger et al., Nature 314: 268-270 (1985); Taniguchi et al., EP-A-171496; Morrion et al., EP-A-173494; Neuberger et al., WO 86/01533; Kudo et al., EP-A-184187; Sahagan et al., J. Immunol. 137: 1066-1074 (1986); Robinson et al., WO

5   87/02671; Liu et al., Proc. Natl. Acad. Sci USA 84:3439-3443 (1987); Sun et al., Proc. Natl. Acad. Sci USA 84:214218 (1987); Better et al., Science 240: 1041-1043 (1988) und Harlow und Lane, Antikörper: A Laboratory Manual, as quoted above). These references are also included in the present invention, as if disclosed in their entirety.

10

An "anti-idiotypic antibody" according to the present invention is an antibody that recognizes a determinant, which is generally associated with the antigen binding site of an antibody according to the present invention, e.g. an anti-leptin-binding protein antibody. An anti-idiotypic antibody can be produced through immuniza-

15   tion of an animal of the same species and the same genetic type (e.g. a mice strain) as a point of origin for a monoclonal antibody, against which an anti-idiotypic antibody according to this invention is targeted. The immunized animal will recognize the idiotypic determinants of the immunizing antibody through the production of an antibody, that is directed to the idiotypic determinants (namely against an anti-

20   idiotypic antibody according to the present invention), (U.S. 4,699,880). An anti-idiotypic antibody according to the present invention may also be used as an immunogen, in order to provoke an immune response in another animal and to induce the production of a so-called anti-anti-idiotypic antibody there. The anti-anti-idiotypic antibody may be, but does not have to be, identical to the original mono-

25   clonal antibody with reference to the design of its epitope, that had caused the anti-idiotypic reaction. This allows the identification of other clones, that express antibodies of identical specificity, with the use of an antibody directed to idiotypic determinants of a monoclonal antibody.

12

In order to induce binding of anti-idiotypic antibodies in the respective animals, such as e.g. the BALB/c mouse, monoclonal antibodies, directed to a physiological binding protein of a physiological ligand,e.g. leptin-binding protein, solubilized or suspended in body liquids, can be used. Cells taken from the spleen of such an im-

5   munized mouse can be used to produce anti-idiotypic hybridoma-cell lines, that secrete anti-idiotypic monoclonal antibodies. Furthermore, anti-idiotypic mono-clonal antibodies may also be coupled to a medium (KLH, "keyhole limpet hemo-cyanin") and subsequently be used for further immunization of BALB/c-mice. The sera of these mice contain anti-anti-idiotypic antibodies, that exhibit the binding

10   properties of the original monoclonal antibodies and that are specific for a physiol-ogic binding protein solubilized or suspended in body liquids of a physiologic ligand (ref. preferred examples below). Therefore, the anti-idiotypic monoclonal antibodies have their own idiotypic epitopes or "idiotopes", characterized by a simi-lar structure as the structure of the epitope to be examined

15

In a preferred embodiment of the invention, the inventive antibody is directed to the extra-cellular domain of a leptin receptor, in particular to a leptin-binding protein. In a more preferred embodiment, the antibody A according to the present invention is directed to the leptin-binding site on a leptin-binding protein, to which the ligand

20   (e.g. leptin) binds. Furthermore, the ligand is preferably leptin. In this context, the term "antibody directed to the leptin-binding site on a leptin-binding protein", means that the antibody as a binding epitope binds specifically and with high affin-ity to the binding site of the ligand (e.g. leptin) on the leptin-binding protein.

25   Furthermore, an antibody of the present invention may also be bispecific, that is to say, it may also recognize different epitopes with its two paratopes, preferably two different epitopes of the same protein or peptide (see above). Eventually, both para-topes may be structural different, however, they may still bind the same epitope or at least overlapping areas of these epitopes.

30

13

According to another embodiment, the antibody A according to the present invention is humanized and directed to a human leptin receptor or a human leptin-binding protein. Humanization of antibodies is known in the prior art can be performed by a large variety of standard methods. Therefore, human or humanized

5   antibodies are also understood as antibodies according to the present invention.

"Fragments" of an antibody according to the present application are also encompassed by the present invention. A "fragment of an antibody according to the present application" typically may comprise any fragment of an antibody of the present

10   application, either fragments of a polyclonal or monoclonal antibody. A fragment of an inventive antibody thus may comprise e. g. the constant regions of the heavy chain of the inventive antibody, e.g. C_H1, C_H2 or C_H3, the variable heavy (V_H) immunoglobulin domain, the variable light immunoglobulin domain (V_L), or the constant light immunoglobulin domain (C_L). The constant heavy immunoglobulin do-

15   main is typically an F_c fragment comprising the C_H3 domain and/or the C_H2 and/or the C_H1 domain. The variable light immunoglobulin domain is preferably an F_{ab} fragment comprising the V_L domain. Also encompassed are all shortened or modified antibody fragments presenting one or two binding sites complementary to the antigen, such as antibody parts with a binding site corresponding to the antibody,

20   composed of a light and a heavy chain, such as F_v-, F_{ab}- or $F_{(ab}')_2$-fragments or single-chain antibody fragments (scF_v). Shortened double strand fragments, such as F_v-, F_{ab}- or $F_{(ab}')_2$ are preferred. F_{ab} and $F_{(ab}')_2$-fragments have no F_c-fragment, which would be present for instance in an intact antibody, therefore, they may be transported faster in the blood circulation and show comparably less non-specific tissue binding than

25   intact antibodies. In this context, it is stressed, that F_{ab} and $F_{(ab}')_2$ fragments of antibodies according to the present invention can be used in an inventive method in the sense of the invention presented. Such fragments are typically produced by proteolytic cleavage, using enzymes, such as e.g. papain (for the production of F_{ab}-fragments) or pepsin (for the production of $F_{(ab}')_2$, fragments), or by chemical oxida-

30   tion or by genetic manipulation of the antibody genes.

14

Furthermore, fragments of the antibodies of the present invention are typically functionally homolog to the antibodies of the present invention.

5   "Functionally homolog" in the meaning of the present invention means that a fragment, a variant, etc. of an antibody of the present invention preferably recognizes specifically a sequence of a leptin–R or a leptin-BP. More preferably, a functional homolog of an antibody of the present invention recognizes specifically an epitope of a leptin-R or a leptin-BP. Even more preferably, the functional homolog of an antibody of the present invention recognizes specifically the leptin binding site of leptin-R or leptin-BP. A functional homolog of an antibody of the present invention means that this homolog is capable of displacing Leptin from its binding to leptin-R or leptin-BP.

15   A "functional homolog" of an antibody of the present invention is also understood to include antibodies with increased or lowered affinity to leptin-R or leptin-BP and/or increased lowered capability of displacing Leptin from its binding to leptin-R or leptin-BP as compared with a full-length antibody of the present invention. Such a high affinity antibody of the present invention preferably comprises or consists of an amino acid sequence, being encoded by SEQ ID NO: 5 or comprises the amino acid sequence SEQ ID NO: 6. Such antibodies with a modulated activity may excite different biological properties. Furthermore, a person skilled in the art may select an antibody with a specific affinity as necessary in the respective case.

25   In a more preferred embodiment a fragment of an antibody of the present invention comprises or consists of a protein sequence encoded by a nucleic acid sequence selected from SEQ ID NOs: 2, 4, 5, or 7. Alternatively, such a fragment comprises or consists of a protein sequence selected from SEQ ID NOs: 1, 2, 3, 4, 6 or 8.

15

In a specific embodiment, the antibody A according to the present invention is the antibody ZMC2. ZMC2 is a monoclonal antibody, directed to the binding site of human leptin on the human leptin-binding protein and in the framework of the invention it had been optimized for the solution of this task. The monclonal antibody

5   ZMC2 has been deposited in viable form by the applicant with entry dated September 25, 2003 under deposit number DSM ACC 2618 along with the reference "ZMC2", assigned by the depositor, according to the stipulations of the Budapest Treaty at DSMZ (Deutsche Sammlung für Mikroorganismen und Zellkulturen GmbH) in Braunschweig.

10

"Variants" of any of the above mentioned inventive antibodies, particularly variants of the inventive antibody ZMC2, are also contemplated by the present invention. A variant of an antibody, particularly a variant of ZMC2, in the meaning of the present invention typically comprises a sequence, wherein at least one, two or more amino

15   acids, preferably 1-5, 1-10, 1-15 or 1-20 amino acids, of the entire amino acid sequence of said antibody are altered, i.e. deleted, substituted or added with respect to the amino acid sequence of the full-length antibody of the present invention, e.g. ZMC2. Variants of antibodies of the present invention, particularly variants of ZMC2, are preferably functionally homolog to the full-length non-altered antibodies

20   of the present invention, particularly to ZMC2.

"Fusion proteins" are also provided by the present invention. A "fusion protein" according to the present invention typically comprises a portion I and a portion II, wherein portion I may be any antibody or antibody fragment according to the pre-

25   sent invention fused to other (protein)-constituents as portion II. Typically, portion I of such a fusion protein represents a full-length antibody. Alternatively, portion I of such a fusion protein may comprise any of the above mentioned fragments of an inventive antibody, preferably V_H or V_L. (Protein)-constituents as portion II of a fusion protein of the present invention typically comprise any antibody or antibody

30   fragment according to the present invention, or any peptide or protein, forming in-

16

teractions with other proteins, e.g. ligands of leptin-R and/or leptin BP, preferably leptin or fragments or variants of leptin. It is preferred to use components as portion II, which specifically recognize cells to be targeted by the inventive fusion protein. Cells to be targeted are typically adipocytes, connective tissue cells and other leptin

5   binding cells. A fragment of a leptin as used in the inventive fusion protein is preferably a shortened leptin molecule, still capable of binding to the leptin binding site of leptin-R or leptin-BP. A variant of a leptin as used in the inventive fusion protein is preferably a full-length leptin molecule, comprising one or more amino acid substitutions , but still being capable of binding to the leptin binding site of leptin-R or

10   leptin-BP. Therefore, the inventive fusion protein is preferably "bispecific", i.e. it is capable of binding two target molecules, whereby portion I is specific for target molecule I and portion II is specific for target molecule II. If the inventive fusion protein comprises e.g. leptin as portion II of the fusion protein, it may act as a so called "super-antagonist". This is due to the fact that leptin binds as a dimer to. Conse-

15   quently, a fusion protein consisting of an inventive antibody (or a fragment thereof, both acting as an antagonist) as portion I and leptin (or a fragment thereof, or a high affinity leptin molecule) as portion II enable a fusion protein to bind with a higher strength and a better specificity to leptin-R or leptin-BP and to displace leptin from its binding to leptin-R or leptin-BP even more efficiently than an inventive antibody

20   as such.

Inventive antibodies as constituents of fusion proteins (as portion I or II) are typically provided by using the methods of enzymatic cleavage, protein-synthesis or by using those recombinant methods, as known by the expert in biochemistry or molecular

25   biology.

The inventive fusion protein may comprise an inventive antibody (or a fragment thereof, both acting as an antagonist) as portion I and a second antibody as portion II. Alternatively, the inventive fusion protein may comprise an inventive antibody

30   (or a fragment thereof, both acting as an antagonist) as portion I and a scFv-fragment

of the inventive antibody as portion II. Such a fusion protein allows to potentially target the antibody to a specific cell type e.g. to obtain bispecific fusion proteins thereby. Preferably, the bispecific fusion protein is directed to a leptin receptor and/or a leptin-binding protein as first specificity and to a cell surface protein as

5 second specificity of portion II. The inventive antibody, such as ZMC2, may be linked to an antibody, such as CD4-F_{ab}, directed to e.g. CD4 (or CD1, 2, 3, 4, 5, 6, 7, 8, 9, 10, CD 25, CD44, etc.). E.g. antibodies recognizing cell surface proteins of specific immune cells allow an inventive fusion protein to specifically bind to immune cells. In this case the affinity of both F_{ab}'s could be altered to get a preferential

10 effect. For example, a high affinity CD4-F_{ab} as a bispecific antibody could then target CD4 cells.

Portions I and II of the inventive fusion protein optionally may be linked by a linker sequence. Peferably, the linker comprises a length of about 5 to 40 amino acids,

15 more preferably between 5 and 30 amino acids and most preferably between 5 and 20 amino acids. Also preferably, the linker comprises a sequence containing at least 50% glycine residues, preferably at least 60 %, more preferably at least 70% and most preferably at least 80%.

20 In a specifically preferred embodiment, the fusion protein of the present invention comprises or consists of a protein sequence encoded by the nucleic acid sequence SEQ ID NO: 7 or of the protein sequence SEQ ID NO: 8.

Furthermore, an inventive antibody, a fragment thereof or a fusion protein according

25 to the present invention, may have further covalently (or not covalently) coupled molecules or groups, for example a fluorescence marker or other markers, for example a gold label, or specific epitopes, that can be recognized by third molecules.

The subject matter of the invention disclosed herein are also mixtures (composi-

30 tions) of the inventive antibodies in the above sense, for instance mixtures of mono-

18

clonal antibodies or mixtures of monoclonal antibodies with antibody fragments, mixtures of anti-idiotypic antibodies etc..

5   Another embodiment of the present invention is a method for preferably quantitatively determining a ligand in a sample containing the ligand of a binding protein, e.g. in solubilized or suspended form, wherein at least one antibody A or an inventive fusion protein according to the present invention is added to the sample to be determined (step a). This inventive method (step a) may be followed by a second step (step b) determining the ligand concentration in the sample.

10

An "antibody A" as used for the present inventive method is defined as given above.

According to the present inventive method, the antibody A or an inventive fusion protein may be added to the sample before or during, preferably before, quantitative
15   determination of the ligand concentration, e.g. leptin, in the sample, and/or may be incubated together with the sample.

An antibody A according to the present invention, that binds exactly and with high affinity to the site of the leptin-binding proteins, to which the ligand binds, is espe-
20   cially preferred. Due to the binding of the inventive molecule, e.g. an antibody A or an inventive fusion protein, a displacement of the ligand (leptin) from the leptin-binding protein occurs, preventing simultaneously binding of the ligand. This displacement will obviously be dependent on the increase of the amount of antibody A or inventive fusion protein added. Thus, a major excess of antibody A or inventive
25   fusion protein over the molar leptin-binding protein concentration is mostly favourable for carrying out the inventive method.

The ligand (e.g. leptin) is "released" through the displacement and can subsequently be measured quantitatively without steric inhibition due to the binding activity of
30   the leptin-binding protein.

19

A "ligand" as used in an inventive method typically comprises all compounds, that bind with high affinity to a leptin receptor and/or a leptin-binding protein and allow a measurement of their concentration in specific assays, and all compounds that are

5   also determined, for instance for medical purposes. E.g. messenger substances, such as hormones, transmitters, for instance neurotransmitters, extra-cellular signal pep- tides or proteins, cytokines, chemokines, lymphokines etc. are compounds in the meaning of the present invention. Leptin is a preferable ligand of the invention. A ligand in the meaning o the present invention may also be a "physiological ligand".

10   Such a "physiological ligand" is typically a ligand in the above sense, which is found in the body of a vertebrate, in particular in a body liquid, without being added exogenously. Correspondingly, this is also applicable for the physiological leptin-binding protein, where this is also the leptin-binding protein of the physio- logical ligand, naturally to be found under physiological conditions.

15

A "sample" in the sense of this invention is typically to be understood as any type of solution to be tested, in particular solutions of medically relevant substances, such as e.g. blood, lymph, serum, urine, liquor, also in a processed form, prepared for the sample handling. Likewise, it is preferable, if the sample to be determined in the

20   inventive method contains liquid, preferably body liquid, more preferably human body liquid, in particular blood or human blood.

A "body liquid" is to be understood as each liquid obtained from the body of a ver- tebrate, in particular a mammal, in particular of a human being. In the case of hu-

25   man beings, this would for instance be blood, urine or lymph, but also cytosolic preparations from human cells.

The term "in solubilized or suspended form" in the meaning of the present invention is to be understood as any form of solution of the leptin-binding proteins or of the

30   ligand in the broader sense in the solution medium. This also includes situations, in

20

which for instance the leptin-binding protein is about to precipitate or has already precipitated, as long as it is still a constituent of the solution.

"Quantitative determination" in the context of the inventive method is to be under-
5   stood as any method known by a skilled person for determining the amount of an analyte solved in a sample. This includes explicitly, e.g. quantification by using chromatographic methods with a concurrent standard, in particular a quantification by using the highly affine interaction between antibody and ligand (antigen) via competitive assays or binding assays, such as e.g. a sandwich assay, also in the
10   ELISA format. A specific advantage of this invention is the fact that adding a specific antagonistic antibody or an inventive fusion protein to a sample as disclosed herein can be combined with practically all known – in particular commercially available – test kits or test systems for quantitative analysis of the respective analyte (ligand) without any efforts.

15

In the inventive method preferably the antibody A or an inventive fusion protein is added prior to or during, preferably prior to the quantitative determination and/or is incubated with the sample.

20   The sample to be tested is typically incubated together with antibody A or an inventive fusion protein. The term "incubation", as used herein is to be understood as a reaction condition, in which the reaction partners, in other words antibody A or an inventive fusion protein and leptin-binding protein are allowed to react with each other. Typically, antibody A or an inventive fusion protein is added to a sample con-
25   taining leptin binding protein and leptin. The incubation is generally carried out for a limited period of time, in this case for example 6, 12, 18 or 24 hours. The term incubation has to be understood primarily as prior step – for example for a period of 6, 12, 18 or 24 hours –before the start of the quantitative measurement (for instance with a commercially available test kit) using antibody B for ligand (leptin) detection.

30

In another preferred embodiment of the method according to the present invention, the leptin-binding protein is a physiological partner of the ligand.

The method according to the present invention is also useful, when the ligand
5 and/or the leptin-binding protein has been added externally (exogenously), e.g. following an injection or other uptake of, for instance, leptin as a ligand in patients or test persons.

10

The present inventive method can be used to measure any ligand of "binding proteins". Binding proteins are proteins interacting with other proteins. An example of a binding protein is leptin-binding protein or the leptin-receptor, interacting with leptin. The binding protein, as used in a method according to the present invention,
15 is preferably soluble, more preferably a soluble (leptin-)receptor.

In a preferred embodiment of the method according to the present invention, the ligand as bound by a "binding protein" is a peptidic compound and/or a hormone, preferably a peptidic hormone, in particular leptin, or a hormone-binding protein.
20 The term "peptidic" is to be understood here to comprise any compound, of which constituents are predominantly linked together by a peptidic bond (such as R1-NH-C(O)-R2).

Hormone binding proteins, if used as ligands in the inventive method, are preferably
25 binding proteins (see below), that bind affine hormones. A "hormone" herein is to be understood as a chemical messenger substance, that acts at a distance from its site of synthesis and liberation. Hormones are preferably produced by endocrinal glands, for instance the hypophysis, gonads or epiphysis. Examples are the growth hormone or the luteinizing hormone LTH, insulin, melatonin, glucagon, gastrin, an-

22

giotensin, substance P, interleukines, vasopressin, endorphines, enkephalines, re-
laxin, the atrionatriuretic factor or also leptin.

In a specifically preferred embodiment of the method according to the present in-
5   vention, the ligand used in such a method is human leptin and the leptin-binding
protein is human leptin-binding protein. Furthermore, the antibody A or an inven-
tive fusion protein is preferably directed to the binding site of leptin on the leptin-
binding protein.

10   Moreover, it is preferred to add a further antibody B, preferably a monoclonal anti-
body B, if a ligand of the binding protein according to the inventive method is quan-
titatively determined.   Such an "antibody B", preferably a "monoclonal antibody
B", is typically an anti-ligand-antibody, i.e. an antibody, which is directed to any
ligand according to the present invention but not to leptin-R or leptin-BP. It is ad-
15   vantageous, if quantitative determination in the inventive method is carried out by
using a competitive binding test, preferably a "radio-immuno assay" (RIA). Likewise
it is typically favourable to quantitatively determine the ligand concentration by
reading out a signal, which depends on the concentration of the ligand to be deter-
mined, preferably by using an enzyme-linked immuno sorbent assay (ELISA) and/or
20   an according sandwich-assay.

In another preferred embodiment of the method according to the present invention,
no separation of the leptin-binding proteins from the sample to be determined is
effected prior to quantitative determination. In particular, this means, that it is not
25   required for the invention disclosed to extract the leptin-binding protein from the
sample prior to its quantitative analysis, in contrast to the state-of-the-art methods
for the determination of ligand concentrations. In the art, practically each form of
extraction is carried out, in which the ligand to be determined – such as for instance
the hormone – remains in the sample, while the binding protein is separated from
30   the sample, e.g. by precipitation and/or filtration with determined molecule weight

23

exclusion limits, chromatographic methods, such as e.g. affinity chromatography or
HPLC, dialysis in determined ligand sizes etc. Precisely this step can be avoided,
preferably while using the method according to the present invention. Due to the
selection of antibody A or an inventive fusion protein, neither antibody A/the inven-
5   tive fusion protein nor the complex formed of antibody A/the inventive fusion pro-
tein and leptin-binding protein is able to bind to the ligand to be determined.
Therefore, in the sample determination is no longer interfered or falsified by
ligand/binding protein interaction, even if the complexed leptin-binding protein re-
mains in the sample. However, use of the highly affine antibodies B for quantitative
10   measurement does not interfere, since they do not bind to antibody A/the inventive
fusion protein, to the leptin-binding protein or the complex formed by both, but ex-
clusively to the ligand itself.

This advantage is in particular observed in an especially favourable and preferential
15   method according to the present invention, in which the antibody A/the inventive
fusion protein is added in such a way, that subsequent to addition the molar con-
centration of antibody A is equal or preferably higher than the molar concentration
of the leptin-binding protein, preferably at least 50 % higher, more preferably at
least twice as high, more preferably at least three times as high and in particular at
20   least four times as high as the leptin-binding protein concentration.

Precisely, this addition of the antibody A/the inventive fusion protein in excessive
amounts to the leptin-binding protein is especially favourable, since in the presence
of a sufficient excess of antibody A/the inventive fusion protein all ligands are dis-
25   placed entirely from the leptin-binding protein and therefore become measurable.
Since the leptin-binding protein is quantitatively complexed, it does not interfere
any more with ligand concentration.

The correct and sufficient amount of specific antibody A or of inventive fusion pro-
30   tein to be added to the sample in order to provide a quantitative measurement of the

24

ligand without interference phenomena, can be determined by the expert by simple preliminary tests. The optimal addition depends on the amount of a leptin-binding protein in the sample and eventually on the quantitative test system used or the amount of ligand to be determined.

5

In a preferred embodiment of the present invention a ligand is quantitatively determined by using antibody B, directed to the ligand, preferably a monoclonal antibody B, in a sample containing human blood. Therein, the ligand and a leptin-binding protein are preferably contained in solubilized or suspended form, and an-

10   tibody A/an inventive fusion protein is directed to the binding site of the ligand on the leptin-binding protein. Preferably the deposited monoclonal antibody ZMC2, is added as antibody A to the sample to be determined in significant excessive amounts as compared to the amount of leptin-binding protein, before or during quantitative determination of the ligand, preferably before step b (quantitative de-

15   termination, by adding and incubating antibody B).

Preferably, the ligand as used in a method according to the present invention is leptin.

20   The present inventive method typically prevents interference of the leptin-binding protein with leptin by adding highly excessive amounts of a specific, preferably monoclonal antibody A, or an inventive fusion protein to the sample to be determined. Antibody A or an inventive fusion protein are directed to the leptin-binding protein. An antibody A as used in the inventive method is preferably the mono-

25   clonal antibody ZMC2. It has been selected due to its particularly preferred property to bind to the leptin-binding protein-molecule exactly at the same binding site as the leptin molecule does. If excessive amounts of antibody A are added, it leads to a displacement of the leptin molecules from their binding to the leptin-binding protein. Leptin is "released" and may then be measured by any immunoassay for leptin

30   without sterically induced blocking by leptin-binding protein. Since the "anti-leptin-

25

binding protein-antibody" (antibody A) itself or the inventive fusion protein itself do not interfere with the "anti-leptin antibodies" (antibody B) used for the measurement of leptin (ligand), removal of the complexes consisting of antibody A/inventive fusion protein and leptin-binding protein from the sample is irrelevant. Practically, e.g.

5   special anti-leptin-binding protein antibody ZMC2 with its leptin-displacing property or an inventive fusion protein is added to the sample before the analysis. After incubation with ZMC2 ligand determination follows the respective manufacturer's guidelines.

10 Another embodiment of the invention relates to a medicament or a vaccine containing at least one inventive antibody A and/or at least one inventive fusion protein. Preferably, a medicament or a vaccine is disclosed, which contains at least one antibody A or an inventive fusion protein according to the present invention and optionally additives and/or adjuvants. Eventually, further active substances may be

15 present in the medicament or vaccine according to present invention.

The medicament or vaccine containing at least one antibody A and/or an inventive fusion protein according to the present invention is suitable, for instance for the treatment of excessive leptin levels. The inventive antibody may inhibit - preferably

20 entirely – all leptin receptor binding sites. As a consequence, leptin cannot bind to its leptin receptor and/or the therapeutic antibody A/the inventive fusion protein displaces leptin, which is bound to a leptin-binding protein, from its binding site.

According to a further embodiment, an antibody A/the inventive fusion protein ac-

25 cording to the present invention or a medicament or vaccine containing at least one inventive antibody A/the inventive fusion protein may be used therapeutically as a leptin-antagonist or may be used for the preparation of a medicament, that inhibits the physiological effect of human leptin. The physiologic effect is blocked by an antibody A/the inventive fusion protein according to the present invention due to its

30 binding to the leptin binding site of the leptin-binding receptor.

A medicament or vaccine, containing one or more antibodies A according to the present invention (and, if applicable, further adjuvants or additives) and/or an inventive fusion protein, can therefore be used for therapeutic purposes also in any of

5   those diseases, disorders or conditions showing unphysiologically increased leptin levels. This also implies, according to the present invention, the use of such inventive antibodies A/fusion proteins, medicaments or vaccines containing at least one antibody A for the treatment (or for the preparation of a medicament for the treatment) of all those disorders, diseases or pathophysiologies, for which excessive

10   leptin levels are etiological, for example diseases of the energy metabolism, pathologic eating disorders, such as anorexia or cachexia. The inventive antibodies A or an inventive fusion protein, medicaments or vaccines, containing an antibody A, may also be used in the treatment of TH1 mediated diseases including multiple sclerosis, diabetes, type 1 diabetes, chronic heart failure (CHF), TNF-mediated diseases,

15   autoimmune colitis, rheumatoid arthritis, systemic lupus erythematosus, and transplant rejection, for the regulation of increased proliferation of naturally occurring regulatory/suppressor T cells and treatment of diseases associated therewith, and diseases associated with the MAPK/ERK1-2, AKT, p-27-kip1 signalling pathways.

20   Moreover, there are experimental results that document a correlation of leptin values and autoimmune disorder disposition.

The inventive antibodies A, medicaments or vaccines, containing an antibody A or an inventive fusion protein, may also be used to block immune effects of leptin. The

25   inventive antibodies A, fusion proteins, medicaments or vaccines, containing an antibody A or a fusion protein, therefore may be used for immune therapy.

By administration of at least one antibody A according to the present invention, or an inventive fusion protein or the respective inventive medicaments or vaccines, the

30   effects of pathophysiologically increased extra-cellular leptin-concentration can be

27

blocked without decreasing its serum concentration. Therefore, it is highly useful and promising, to use the inventive medicament or vaccine also for the treatment of diseases such as anorexia nervosa and in the different stages of cachexia. The hyper-secretion of leptin in the case of insulin-dependent diabetes mellitus, which is made
5   responsible for a variety of sequelae, may also be treated by an leptin-antagonist according to the present invention or a medicament or vaccine containing at least one antibody A.

Where appropriate, a therapeutic treatment of affected patients with excessive leptin
10   levels may carried out in combination with such medicaments or active substances, that decrease the secretion of leptin.

A medicament or a vaccine according to the present invention, an inventive anti-body A or an inventive fusion protein is particularly preferred or may serve for the
15   preparation of a medicament, for the treatment of conditions, that are associated with an undesired activation of the immune system as well as with autoimmune dis-eases. Autoimmune diseases may be selected e.g. from multiple sclerosis (MS), rheumatoide arthritis, diabetes, diabetes type I, systemic lupus erythematosus (SLE), chronic polyarthritis, Basedow's disease, autoimmune forms of chronic hepatitis,
20   colitis ulcerosa, allergie type I-diseases, allergie type II-diseases, allergie type III-diseases, allergie type IV-diseases, fibromyalgie, alopecia, Morbus Bechterew, Mor-bus Crohn, Myasthenia gravis, neurodermitis, polymyalgia rheumatica, progressive systemic sclerosis (PSS), psoriasis, Reiter-syndrome, rheumatic arthritis, vaskulitis, etc..

25
In a specific embodiment, a medicament according to the present invention, an in-ventive antibody A or an inventive fusion protein may be used or may serve for blocking peripheral effects of leptin. "Peripheral" effects are evoked by inhibiting leptin effects indirectly via blocking the leptin receptor. In contast, "central" inhibi-
30   tion of leptin are due to direct interaction of a therapeutic compound with leptin.

28

Leptin acts peripherically e.g. on the immune system, whereas central effects are based on its binding to central control organs, e.g. the hypothalamus. Leptin's binding to the hypothalamus leeds to weight control effects. For the treatment of certain immune disorders the interaction of leptin with its receptor shall be inhibited,

5   whereas the leptin level in the other body parts (e.g. hypothalamus) shall remain unchanged in order to avoid considered weight loss/weight gain.

State-of-the-art treatment of leptin excess e.g. by administering anti-leptin-antibodies, lowers the leptin level in the body. Upon administration if anti-leptin-

10   antibodies positive effects on the patient's immune disorder may be achieved. However, simultaneous side effects may appear, such as weight gain due to decreased levels of free leptin, e.g. at the hypothalamus. The inventive antagonist allows to avoid the side effects induced by state-of-the art anti-leptin-antibodies. The anti-leptin-receptor antibody therapy according to the present invention as disclosed

15   herein does not reduce the patient's leptin level by selectively blocking the leptin receptors of certain tissues without modifying the leptin responsiveness of other tissues ("central" effect in the hypothalamus). By way of example, while using an inventive antibody A, such as ZMC2, one might prevent an immune response without having an effect on weight, i.e. there is no significant change in weight during

20   treatment with the inventive antibody A, e.g. ZMC2, or the inventive fusion protein.

Typically, an inventive antibody A or an inventive fusion protein as used in a medicament or a vaccine according to the present invention will be available as lyophilized powder, containing between 0.5 mg and 100 mg of the inventive anti-

25   body A, and further additives, as for example glycin, manitol, and/or sodium phosphate monohydrate. This lyophilized powder is provided in a suitable aqueous solution and administered subsequently, for example subcutaneously once or several times per day.

29

In principle, the medicaments according to the present invention may also be administered as liquid dosage form, in particular in form of injection solutions. Suitable additives and/or adjuvants are e.g. solutions or diluents, stabilizers, suspension mediators, buffer substances, preservatives, as well as colorants, expanders, and/or

5   binders. The selection of the adjuvants as well as of the amounts to be administered depends on whether the medicament shall be administered parenterally, intravasally, intravenously, intraperitoneal or intramuscular. Preparations in form of suspensions and solutions as well as dry preparations, which allow an easy reconstitution, are suitable for all the parenteral applications.

10

Another embodiment of the invention is a diagnostic agent containing at least one antibody or fusion protein according to the present invention as well as, where appropriate, additives and/or adjuvants. A "diagnostic agent" is to be understood as a preparation or an adjuvant which may be useful, for example for the diagnosis of a

15   determined disease.

A further embodiment of the invention is a kit containing distinct from each other at least one first preparation containing at least one antibody A or fusion protein according to the present invention and one operating test assay ready for use, based

20   on an antigen/antibody-reaction, for the quantitative determination of a ligand, that serves as an antigen in this test.

A kit is to be understood as a conjunct form of different constituents in a packaging form. In this case it is in particular a diagnostic-kit, that contains the different con-

25   stituents required for the quantitative analysis of a ligand.

A kit is preferred, in which the first preparation contains an antibody A or fusion protein, directed to the binding site of the ligand, preferably leptin, especially preferentially human leptin, on the, preferably human, leptin-binding protein, and/or

30   which contains, apart from the first preparation and the operative test assay ready

30

for use, based on an antigen/antibody-reaction, for the quantitative determination of a ligand, also a preparation for the calibration.

An especially preferable embodiment is a kit according to the present invention, in
5   which the first preparation contains the antibody ZMC2 (deposited, see above) and/or the preparation for the calibration and/or the antibody B in the enclosed operating test assay ready for use, wherein the test assay is based on an antigen/antibody-reaction for quantitative determination of a ligand, wherein antibody B, preferably a monoclonal antibody B, is directed to the ligand.
10
Preferably, the ligand is leptin and the antibody B is directed to the main isoform of leptin with a molecular weight of 16 kDa.

A further embodiment of the invention is the use of at least one antibody for deter-
15   mination, preferably quantitative determination, of a physiological ligand, preferably leptin, of a physiological leptin-binding protein.

A further subject matter of the invention is a method for preparing an antibody ac-
20   cording to the present invention, i.e. an antibody directed to a leptin-binding protein, which is further directed to a ligand, comprising the following steps: (a) immunization of animals with recombinantly produced leptin-binding protein, (b) isolation of immune cells from the animal, (c) fusion with myeloma-cell lines to hybridoma cell cultures, (d) selection of clones with high specificity for the leptin-binding
25   protein. The immunization can carried out with all animals suitable for such purposes, such as mice, rabbits, pigs, horses etc. In order to separate such clones, that produce antibodies directed to a leptin-binding protein, from inventive clones (that produce antibodies, which are e.g. specifically directed to the ligand binding site of the leptin-binding protein), suitable media (e.g. wells of microtiter plates, polystyrol
30   beads, plastic tubes) are each coated with one (unselected) anti-leptin-capture anti-

31

body in a method step (e). In another method method (f), the leptin-binding protein and subsequently the ligand (e.g. leptin) are added. Thereby, the ligand should is preferably labeled (for example by using biotinylation, label (radioactive label, fluorescence marker, enzyme marker (horseradish-peroxidase) etc.)) or be detectable by using a respective anti-ligand-antibody, such that the property of the ligand, to bind to the leptin-binding protein in the recipient coated with capture antibody can be examined. Thus, alternative methods are for instance radioactivity, chemoluminescence, colorimetric methods or an enzyme reaction. After the washing step, those "wells" or "beads" that do not show a signal after the addition of the ligand, can be identified. In this case, the capture antibody blocks the ligand binding site. Thus, the binding site is not available any more for the binding of the ligand, since the capture antibody interferes with the ligand.

Finally, an antibody A or a fusion protein according to the present invention can be identified with by carrying out competitive binding assays. As described above, a coating process is required here, though, in this method each well (e.g. of a microtiter plate) is coated with an anti-leptin-binding protein-antibody, that does not specifically recognize the ligand binding site. After addition of the leptin-binding protein, a potentially interesting antibody or a fusion protein (each with the property of specifically recognizing the ligand binding site) is added to each well above with labeled ligand. As the concentration of the antibody specific for the ligand binding site increases, the signal intensity of the ligand in the well decreases. This method may be used for selection of inventive antibodies directed to a leptin receptor and/or a leptin-binding protein.

As an alternative to using hybridoma cells, so-called "phage display" methods (Morphosys or Cambridge Antibody Technologies) may be used, in order to generate potentially inventive antibodies and to select them, as described above.

32

In the following section, the invention is described in a more detailed way by ex-
amples, without limiting the scope of the present invention to these Examples.

5   Examples and Figures:

Description of Figures:

Figure 1:   shows the results of the examination in Example 2. The results prove,
10   that addition of the inventive antibody may diminish or abolish the in-
             fluence, which leptin mediates via the leptin receptor.

Figure 2A: shows the protein sequence of the ZMC2/F_{ab} light chain (amended).

15 Figure 2B: depicts the nucleotide sequence of the open reading frame of the
                  pComb3/ZMC2,F_{ab} vector (light chain) and its translation into the en-
                  coding protein sequence of ZMC2/F_{ab} light chain (cf. also Figure 2A).
                  Start and stop codons are indicated in bold letters, restriction sites are
                  underlined, and primers sequences are double underlined. The se-
20   quence of the pComb3/ZMC2,F_{ab} vector is indicated in italic letters.

Figure 3A: depicts the protein sequence of ZMC2/F_{ab} Heavy chain-His.

Figure 3B: shows the nucleotide sequence of the open reading frame of the
25   pComb3/ZMC2,F_{ab}-His vector (heavy chain) and its translation into the
              encoding protein sequence of the heavy chain of ZMC2/F_{ab} Heavy
              chain-His (cf. Figure 3A). Start and stop codons are indicated in bold
              letters, Restriction sites are underlined, and primers sequences are
              double underlined. The sequence of pComb3/ZMC2,F_{ab}-His vector is

indicated in italic letters. The His-tag is indicated by a dotted (..........)
line.

Figure 4A: discloses the nucleotide sequence of the high affinity clone ZMC2
ScFv (1C3). The V_L sequence is indicated in bold letters. The V_H sequence is double underlined. The linker in between is indicated in normal letters. The sequence of the vector is indicated in italic letters. The E-tag is indicated by a dotted line. Restriction sites (SfiI and NotI) are underlined, and primers sequences are double underlined.

Figure 4B: shows the translated protein sequence encoded by the nucleotide sequence of the high affinity clone ZMC2 ScFv (1C3) as shown in Figure 4A. Disclosed is the sequence from V_H until the end of the E-tag.

Figure 5A: depicts the nucleotide sequence of the ZMC2 Heavy chain – Leptin construct. Disclosed is the 1203 bp nucleotide sequence, wherein start and stop codons are indicated in bold letters and restriction sites are underlined. The ZMC2 heavy chain is indicated by a dotted (.........) line, the $(G_4S)_4$ linker is double underlined. The sequence of Leptin indicated by a (_____) line and is the vector is indicated in italic letters. The his tag is indicated by a (_____) line.

Figure 5B: shows the translated protein sequence of the ZMC2 Heavy chain – Leptin construct (cf. Figure 5A). The ZMC2 heavy chain is indicated by a dotted (...........) line, the $(G_4S)_4$ linker is double underlined. The sequence of Leptin indicated by a single line and is the vector is indicated in italic letters. The his tag is indicated by a (_____) line.

34

Figure 6:   illustrates the binding of ZMC2 antibody to human leptin-R on ELISA. Figure 6 clearly shows that ZMC2 antibody binds to human leptin-R in a dose dependent manner (cf. Example 3).

5   Figure 7:   shows a bar plot indicating the results of a competitive binding experiment of a ZMC2 antibody to human leptin-R in the presence of (excess) Leptin. On the x-axis different concentrations of leptin (ng/ml) are indicated (10000, 1000, 100, 10, 1, and 0,1 ng/ml) and ZMC2 in a concentration of 10 ng/ml). The y-axis shows the absorbance at 450

10   nm (OD 450 nm). As can be seen, the binding of ZMC2 to leptin-R is apparently very strong. Only a large excess of leptin can displace the binding of ZMC2 antibody to human leptin-R (cf. Example 4).

Figure 8:   shows a bar plot of the dose response of luciferase activity in response

15   to leptin stimulation in HEK293 cells. Therefore, HEK293 cells were transiently transfected with Ob-Rb and a STAT3-luciferase reporter construct and stimulated with varying doses of leptin for 2 hours. Then, luciferase activity was measured and was corrected for B-galactosidase. On the x-axis different concentrations of leptin (ng/ml)

20   are indicated (1, 5, 10, 20, 50, 100, 250, 1000 ng/ml). The y-axis shows the degree of fold induction versus unstimulated cells. As can be obtained from Figure 8, induction of HEK293 cells with leptin reaches a maximum level, whereby nearly no further stimulation is observed (cf. Example 5).

25
Figure 9:   illustrates in a bar plot the dose response for inhibitory effects of ZMC2 antibody on leptin signalling. HEK293 cells were transiently transfected with leptin-R and a STAT3-luciferase reporter construct and stimulated with leptin (5ng/ml) for 2 hours. Different doses of

30   ZMC2 antibody (ug/ml, see x-axis) were added to the culture,

35

luciferase activity was measured and corrected for B-galactosidase. The results are expressed as fold induction over the unstimulated cells. On the x-axis different concentrations of leptin (ng/ml) (0, 12,5 and 100 ng/ml) and ZMC2 (10, 5, 1, 0,5, and 0,1 ng/ml) are indicated. As can be obtained from Figure 9, ZMC2 antibody demonstrated inhibitory effects on leptin signalling in a dose response manner (cf. Example 6).

Figure 10:   shows a bar plot of the binding of ZMC2 antibody to mouse leptin-R. Binding of ZMC2 antibody was detected using a biotinylated secondary antibody followed by SAV-HRP. The OD of the resulted colour was measured at 450nm. On the x-axis different concentrations of leptin (ng/ml) and ZMC2 (ng/ml) are indicated. As may bee seen in Figure 10, ZMC2 antibody binds to mouse leptin-R in a dose dependent manner and its binding can be displaced by leptin at high concentrations of leptin only (cf. Example 7).

Figure 11:   illustrates in a bar plot the blocking of TNF-α production in leptin activated human monocytes by ZMC2 antibody. Human monocytes were cultured in the presence of PBS, Leptin (250 ng/ml), ZMC2 (10 ng/ml) + Leptin and ZMC2 (5 ng/ml) + Leptin. The Y axis shows percentage of monocytes expressing TNF-α (cf. Example 8).

Figure 12A   depicts a proliferation assay showing the stimulation by okt3 (optimal dose 100ng/ml) of human PBL in autologous human serum in the presence of ZMC2 ab in different concentrations. In Figure 12A ZMC2 is shown (from 001, 0,1, 1 and 10 ug/ml) in the presence or absence of exogenous leptin (100ng/ml). As may be obtained from Figure 12A, there is a minor effect of ZMC2 to increase PBL proliferation that is reduced by leptin (cf. Example 9).

Figure 12 B   shows the stimulation by okt3 (optimal dose 100ng/ml) of human PBL in autologous human serum with different doses of leptin and more points on ZMC2 mAb dose response (0, 0.005, 0.05, 0.5, 5, 50 ug/ml) are shown. The curve in Figure 12B, is biphasic with a slight tendency to increased proliferation after treatment with ZMC2 which is again reduced by leptin (cf. Example 9).

Figures 12C-D: show comparative experiments carried out with a donor (in the same assay as shown for Figures 12A and B) with an agonistic anti-leptin receptor ab from R&D systems and another anti-human leptin treatment from R&Dsystems. The agonistic anti-ObR2 antibody increases proliferation in a similar fashion to the antagonistic ZMC2 and the anti-leptin at low doses increases the proliferation. The proliferation then is reversed by leptin when using ZMC2, whereas it is still stimulated with anti-ObR2. On a mixed population of naive/memory cells; with leptin present in serum controlling memory cell proliferation, the addition of the blocking ab ZMC2 thus slightly increases proliferation which can then be reversed. Thus, ZMC2 is acting as an antagonistic antibody (cf. Example 9).

Figure 13:   shows a plot of the results from a proliferation assay similar to those as disclosed in Figure 12, but extended to a longer time period (proliferation at 4 days (96 h)). Figure 13 shows the stimulation by okt3 (optimal dose 100ng/ml) of human PBL in autologous human serum with different doses of leptin and more points on ZMC2 mAb dose response (0, 0.005, 0.05, 0.5, 5, 50 ug/ml) are shown. ZMC2 antibody stimulates proliferation, which is then clearly reversed by addition of leptin. Thus, ZMC2 clearly reacts as an antagonist to leptin-R regarding the overall response on the mixed T-cell population (cf. Example 10).

Figures 14A-C: show a plot of food uptake experiments (cf. Example 11) on ob/ob
and ob/+ mice: 3 mice groups were selected; all females; The mice
were initially treated with low dose leptin such as to keep the weight
constant. The treatment was 100ug/daily ip+Rec leptin 0.5ug/gr
weight, for 5 days. In Figure 14A, percent in body weight change is
flat during treatment with leptin alone, and instead of being prevented
by ZMC2, the weight was slightly increased in terms of no change or
even little reduction in body weight. On the heterozygotes (Figure
14B, cf. Example 11), treated in parallel, basically show the similar re-
sults. With the same dose of ZMC2 alone, the untreated mice slightly
increase body weight over the time; In a comparative Experiment (Fig-
ure 14C, cf. Example 9) proliferation of spleen cells from normal
mouse (B6) was stimulated with anti-CD3 for mouse (called 2C11)
(0,0001, 0,001, 0,01, 0,1, 1, 10, 100 ug/ml) dose response in the pres-
ence of FCS 2%. As can be seen in Figure 14C, a clear inhibition can
be obtained at 10-100 ug/ml.

Figure 15:   displays a photograph of a gelelectrophoresis. The samples applied on
the gel were obtained from an anti-CD3 stimulation experiment using
T-cells from autoimmunity-prone NOD mice.   As may be obtained
from the gel ERK1/2 phosphorylation is reduced by the leptin-R block-
ing antibody.

Figure 16:   shows a plasmid map of the pComb3/ZMC2 F_{ab}-His clone.

Figure 17:   shows the results of a Western Blot experiment during purification of
recombinant F_{ab} ZMC2. As can be obtained from the Western Blot,
recombinant F_{ab} ZMC2 is expressed in the soluble fraction and the Blot
shows a protein of the correct size of Kappa light chain.

38

Figure 18:   shows the results of a Western Blot experiment during purification of recombinant His-tagged F_{ab} ZMC2. As can be obtained from the Western Blot, His tagged F_{ab} ZMC2 can be purified on a Cobalt Column.

5

Figure 19:   shows a Coomassie Blue staining of a polyacrylamide gel of a purified F_{ab} ZMC2 antibody.

Figure 20A: shows a bar plot of a binding experiment of ZMC2 and recombinant

10   expressed F_{ab} ZMC2 to human Leptin. The y-axis shows the absorbance at 450 nm (OD 450 nm).

Figure 20B: shows the effect of recombinant F_{ab} ZMC2 on leptin-induced SIE activation. As can be obtained from Figure 20A recombinant F_{ab} ZMC2

15   (rF_{ab}) and chemically made F_{ab} ZMC2 (cF_{ab}) for ZMC2 block leptin signalling.

20

39

Examples

Example 1:

Production of antibody A (directed to leptin-BP)

5

More than 20 Balb/c mice were repeatedly immunized with leptin-BP produced recombinantly according to the generally known method (ref. to leaflet of Titer-max®). When a high titer against leptin-BP was achieved, the spleen has been re-moved from the animals, and according to the method described by Köhler and Mil-

10   stein (Continuous cultures of fused cells secreting antibody of predefined specificity, Nature, 1975, Aug. 7; 256 (5517): 495-7) hybridoma cell cultures were produced by fusion with a mouse-myelom-cell line. Also according to generally known methods (limited dilution, screening of the hybridoma cell culture supernatants with labeled antigen) those clones were selected, which produced monoclonal antibody with

15   high affinity and specificity for leptin- BP.

The majority of the selected clones were directed to such epitopes on the leptin-BP molecule, that are situated externally of the binding site for leptin. In order to select antibodies A according to the present invention, directed to an epitope within the

20   binding site, at first one of many antibodies, that binds leptin-BP externally of the binding site, was used. It was identified by applying all monoclonal antibodies against leptin-BP (as many as possible), e.g. in separate wells each, on a microtiter plate (polystyrol plates with highly adsorptive surface) as capture antibody, (alterna-tively, other methods, as e.g. coated polystyrol beads, coated plastic tubes etc., may

25   also be used). Subsequently, recombinant leptin-BP has been added, which binds to the coating antibody – in a primarily still unknown arrangement (binding according to each epitope within or externally of the interaction site of the hormone receptor). Finally, labeled (in our case biotinylized) leptin has been added, which may now bind exclusively to the leptin-BP-molecules, and which possess a freely accessible

30   binding site (i.e., where the coating antibody does not interfere with the leptin bind-

ing). Determination of bound antibody was carried out by addition of streptavidin-europium, which binds to an bound biotinylized leptin and can be measured in a fluorimeter (time resolved fluorescence after addition of enhancement solution). Alternatively, this step may also be effected using analog methods (radioactivity, en-

5   zyme reaction/colorimetric method or chemoluminescence).

In order to find an antibody, that is directed to the binding site, a microtiter plate was coated with an antibody against leptin-BP, selected according to the above method, that does not interfere with the binding of leptin. The addition of leptin-BP

10   provoked an aligned binding of the leptin-BP-molecules, that is to say with a freely accessible binding site for leptin. Subsequently, biotinylized leptin was added and simultaneously, all other monoclonal antibodies against leptin-BP (once again each antibody in an own well of the microtiter plate). If an antibody is then directed to the binding site, it should block the binding of biotinylized leptin in a concentration

15   dependent manner. This could be detected by a decrease of the signal intensity (detection method in our case again time resolved fluorescence (streptavidin-europium). For confirmation in an inverted experiment a displacement of biotinylized antibodies directed to leptin-BP by adding unlabeled leptin was carried out. This is possible, if the biotinylized antibody competes with the non-labeled leptin

20   for the binding on the receptor binding site, i.e. if the epitope of the mAbs is located at the interaction site of the leptin/ leptin receptor.

Example 2:
Verification of the effectiveness of an antibody according to the present invention

25   and the associated improvement of general quantitative measurement methods in the presence of interfering leptin-binding protein

Leptin-binding protein was immobilized on a microtiter plate such, that the binding site for leptin was freely accessible. This is carried out by using other antibodies di-

30   rected to the leptin-binding protein, that do not bind within, but externally at the hormone-receptor-interaction site. Subsequently, labeled (biotinylized) leptin was

41

added. Then, buffer solutions with ascending concentrations of the inventive anti-body ZMC2 are added to the incubation solutions. Following an incubation time of 2 hours, the amount of labeled leptin bound to immobilized leptin-binding protein was measured by addition of a fluorescence dye, that binds specifically to biotin.

5   With an ascending concentration of ZMC2, the binding of biotinylized leptin de-creased, thus proving the displacement effect of ZMC2.

Furthermore, the functional blockade of the leptin receptors was proven in a cell culture assay by using ZMC2. Therefor, HEK293-cells were transfected transiently

10   with the human leptin receptor as well as with a STAT 3-luciferase-reporter. This construct enables the visualization or the measurement of the intracellular signal cascade activation caused by leptin.

As shown in Figure 1, the addition of leptin to the cell culture provokes a 7-8-fold

15   increase of the luciferase activity (compared with the control containing no leptin). Adding the inventive antibody ZMC2 to the preparation (5 µg/ml) prevents this acti-vation in contrast to the control antibody (antibody with irrelevant specificity, in this case against KLH).

20   Example 3:
Binding of ZMC2 antibody to human leptin-R on ELISA

For detecting binding of ZMC2 antibody to human leptin-R an ELISA plate was coated with recombinant human leptin-R (200ng/well) and different concentrations

25   of ZMC2 antibody (1000, 333, 111, 37, 12,3, 4,1, 1,3, 0,45, 0,15, 0,05, 0,016, 0,005, 0,0018, 0,0006 ng/ml) were added. Binding of ZMC2 antibody to leptin-R was detected subsequently using a biotinylated secondary antibody followed by SAV-HRP and the signal (OD of resulted colour) was measured at 450nm. As a re-sult, ZMC2 antibody binds to human leptin-R in a dose dependent manner as may

30   be obtained from the corresponding bar plot (see Figure 6).

42

Example 4:

Specifity of binding of ZMC2 antibody to human leptin-R

5   For a competitive binding experiment of a ZMC2 antibody to human leptin-R in the
presence of (excess) leptin, an ELISA plate was coated with recombinant human
leptin-R (200ng/well). Different concentrations of leptin (10000, 1000, 100, 10, 1,
0,1 ng/ml) were added to wells in presence or absence of ZMC2 antibody. ZMC2, if
added, was present in a concentration of 10 ng/ml. Binding of ZMC2 antibody was

10   then subsequently detected using a biotinylated secondary antibody followed by
SAV-HRP and the signal (OD of resulted colour) was measured at 450nm. As can be
obtained from Figure 7 only a large excess of leptin can displace the binding of
ZMC2 antibody to human leptin-R. Accordingly, binding of ZMC2 antibody to
leptin-R is highly specific.

15

Example 5:

Dose response of luciferase activity in response to leptin stimulation

In order to detect a dose response of luciferase activity in response to leptin stimula-

20   tion, HEK293 cells were transiently transfected with leptin-Rb and a STAT3-
luciferase reporter construct and stimulated with varying doses of leptin for 2 hours.
After 2 hours the media were replaced with fresh medium containing no leptin and
cultures were inoculated for further 4 hours before the cells were lysed. Subse-
quently, luciferase activity was measured and corrected for ß-galactosidase, which

25   was used as a transfection control. The results are shown in Figure 8. The values
shown express the fold induction over the unstimulated cells. The induction is dose
dependent and reaches a maximum at about 50 – 1000 ng/ml Leptin.

30

43

Example 6:
Dose response for the inhibitory effects of ZMC2 antibody on leptin signalling

To obtain a dose response for the inhibitory effects of ZMC2 antibody on leptin sig-
5   nalling HEK293 cells were transiently transfected with leptin-Rb and a STAT3-
luciferase reporter construct and stimulated with leptin (5ng/ml) for 2 hours. Differ-
ent doses of ZMC2 antibody (10, 5, 1, 0,5 and 0,1 µg/ml, if leptin is contained) were
added to the culture 30 minutes prior to addition of leptin (0 ng/ml, if no ZMC2 is
contained and 12,5 ng/ml, if ZMC2 is contained). Luciferase activity was measured
10   subsequently and was corrected for ß-galactosidase. The results are shown as a bar
plot in Figure 9, being expressed as fold induction over the unstimulated cells. As a
result ZMC2 antibody demonstrated inhibitory effects on leptin signalling in a dose
response manner.

15   Example 7:
Binding of ZMC2 antibody to mouse leptin-R

An ELISA plate was coated with recombinant mouse leptin-R (200ng/well) and dif-
ferent concentrations of ZMC2 antibody, leptin, growth hormone or the combina-
20   tion of them (ng/ml) were added. Binding of ZMC2 antibody was detected using a
biotinylated secondary antibody followed by SAV-HRP and the OD of resulted col-
our was measured at 450nm. The results are shown in Figure 10 as a bar plot. As
can be obtained from the plot ZMC2 antibody binds to mouse leptin-R in a dose
dependent manner and its binding can be displaced by leptin only at high doses.

25

Example 8:
Blocking of TNF-α production in leptin activated human monocytes by ZMC2

For this experiment, TNF-α production was induced by leptin activated human
30   monocytes. After inducing TNF-α production, ZMC2 (10 and 5 ng/ml) was added to

44

the monocytes in the presence and absence of leptin (250 ng/ml).. The results are shown in Figure 11. As can be seen in Figure 11, ZMC2 is capable of blocking TNF-α production by leptin activated human monocytes. Y axis shows percentage of monocytes expressing TNF-α (% double positive (CD-14 and TNF-α positive)

5   monocytes).

Example 9:

Stimulation of human PBL in autologous human serum by okt3 (anti-CD3 experiment)

10

For measuring the stimulation of human PBL in autologous human serum by okt3 (optimal dose 100ng/ml) cells were inoculated in the presence of ZMC2 ab (from 0, 0.1, 1 and 10 ug/ml) and in the presence or absence of exogenous leptin (100ng/ml). Cells were harvested after 60 min allowing maximum DNA synthesis.

15   The proliferation was measured (cpm) and can be seen in Figure 12A. There is a minor effect of ZMC2 to increase PBL proliferation that is reduced by leptin. This increase in proliferation can be reversed by leptin, as can be seen in the following experiments. ZMC2 thus acts as an antagonist. Furthermore, it is considered that the response to anti-CD3 stimulation in humans in the presence of leptin is dependent

20   on the relative proportion of naive/memory cells in PBL; the more naive cells are present the greater is the increase in proliferation in response to leptin; alternatively, the more memory cells are present the greater is the inhibition of proliferation induced by leptin (although IFN-g secretion is increased, despite the reduced proliferation of memory cells).

25

In Figures 12B, C and D stimulation by okt3 (optimal dose 100ng/ml) of human PBL in autologous human serum with different doses of leptin (10 and 100 ng/ml) and more points on ZMC2 mAb dose response (0,0001, 0,001, 0,01, 0,1, 1, 10, 100 ug/ml) is shown. As can be seen, the curve is biphasic with a slight tendency to in-

30   creased proliferation after treatment with ZMC2 (Figure 12B) which is again reduced

45

by leptin; with donor 2 experiments were performed in the same assay, i.e. the ago-
nistic anti-leptin receptor ab (anti-ObR2) from R&D systems (Figure 12 C) and an-
other anti-human leptin treatment from R&Dsystems (Figure 12 D). anti-ObR2 in-
creases proliferation in a similar fashion to ZMC2 and the anti-leptin at low doses

5   increases the proliferation, which is reversed by leptin. Here, the effects on a mixed
population of naive/memory cells are studied, wherein leptin is present in serum
controlling memory cell proliferation. Addition of the blocking ab ZMC2 slightly
increases proliferation, which can be reversed upon addition of leptin. Thus, ZMC2
is acting as an antagonistic antibody, whereas anti-ObR2 acts as an agonistic anti-

10   body.

     Example 10:
     Elongated proliferation experiments

15   To better define the effect of ZMC2, proliferation experiments were carried out over
     a longer time period to see whether the increase in proliferation was more visible
     when cells were to be in a later phase of cell cycle. Therefore, proliferation was
     measured at 4 days (96 h), and surprisingly the ZMC2 ab still stimulated prolifera-
     tion, wherein - looking at the overall response on a mixed T cell population – ZMC2

20   turned out to act as an antagonist, since addition of leptin reduces proliferation (see
     Figure 13). The specificity of ZMC2 additionally can be measured by the fact that
     addition of leptin reduces proliferation.

     Example 11:
25   Food uptake experiments on ob/ob and ob/+ mice: 3 mice group

     For food uptake experiments ob/ob and ob/+ mice, all females, were taken. In case
     of ob/ob, mice were treated with low dose leptin in order to maintain weight, i.e.
     not to loose weight but also not to increase weight, as it usually happens. Usually 1

30   gram of body weight every 2 days is typically fed to the mice. The treatment in the

46

experiment was 100ug/daily ip+Rec leptin 0.5ug/gr weight, for 5 days. As can be seen in Figure 14A the curve in %in body weight change is flat during treatment with leptin alone, indicating that the treatment was working. Instead of being pre-vented by the ab, body weight was slightly increased in terms of no change in body

5   weight. In an comparative Experiment heterozygotous mice (see Figure 14B), were used because they show a lower leptin than normal mice. The heterozygotes were treated in parallel with the same dose of ab alone. The untreated mice showed a tendency to increase body weight over the time (less strong than for the the ob/ob mice); the ab treated were similar for body weight, but after stopping body weight

10   increased more than the controls.

Proliferation of spleen cells from normal mouse (B6)

Proliferation of spleen cells from normal mouse (B6) was stimulated with anti-CD3

15   for mouse (called 2C11) (0,0001, 0,001, 0,01, 0,1, 1, 10, 100 ug/ml) dose response in the presence of FCS 2%. As can be seen in Figure 14C, a clear inhibition can be obtained at 10-100 ug/ml.